

Transformation & Storage Solution (2X TSS)

Cat. No. C905ML

Transformation & Storage Solution (2X TSS) enables researchers to take advantage of the simple system described by Chung *et al.*¹ for the preparation, long-term storage and transformation of competent *E. coli*. Early log-phase cells are suspended in 1X TSS: a solution containing polyethylene glycol, dimethyl sulfoxide, and divalent cations in a bacterial growth medium. The transforming DNA is added to the TSS-treated cells, the mixture is incubated on ice, and the transformed cells are plated on selective media. Transformation efficiencies of 10^5 - 10^6 colonies/ μ g of DNA are routinely obtained without lengthy host cell preparation or heat shock.^{1,2} For example, *E. coli* strains DH1, DH5 α , HB101, JM109, LE392, MM294, SCS-1, and XL-1 gave transformation efficiencies of 1.5 - 6×10^7 colonies/ μ g of DNA. Precise transformation efficiencies will depend on both the host strain and the nature and quality of the transforming DNA.

For added convenience, TSS-treated competent cells can be prepared in advance and stored at -70°C for later use with little or no loss in transformation efficiency. For example, TSS-treated competent *E. coli* JM109 cells showed no significant loss of transformation efficiency after 1-18 weeks at -70°C .¹ However, the transformation efficiency of TSS-treated *E. coli* K802 competent cells was reduced several-fold after similar treatment. Fresh TSS-treated competent cells should be used when the total number of recombinants, the ligation or transformation efficiency or the percentage of correct recombinants is expected to be low.

Product Specifications

TSS is supplied as a 2X solution.

Storage: Store only at -20°C in a freezer without a defrost cycle.

Storage of Competent Cells in TSS: Competent cells frozen in TSS should be stored at -70°C .

Preparation and Transformation of TSS-Competent Cells

Preparation of Competent Cells

1. Dilute an overnight culture of *E. coli* 1:50 with LB broth.
2. Incubate at 37°C with shaking (at 200 rpm) until the cells reach early log phase ($\text{OD}_{600} = 0.25$ - 0.4).
3. While cells are growing, thaw 2X TSS on ice and dilute an appropriate amount 1:1 with sterile distilled water (100 μ l of diluted TSS will be needed for each ml of cells). Chill on ice.

Protocol For Transformation Storage Solution 2x Tss

Sabine Zange



Protocol For Transformation Storage Solution 2x Tss:

cDNA Library Protocols Ian G. Cowell, Caroline A. Austin, 2008-02-02 The first libraries of complementary DNA cDNA clones were constructed in the mid to late 1970s using RNA dependent DNA polymerase reverse transcriptase to convert poly A mRNA into double stranded cDNA suitable for insertion into prokaryotic vectors Since then cDNA technology has become a fundamental tool for the molecular biologist and at the same time some very significant advances have occurred in the methods for constructing and screening cDNA libraries It is not the aim of *cDNA Library Protocols* to give a comprehensive review of all cDNA library based methodologies instead we present a series of up to date protocols that together should give a good grounding of procedures associated with the construction and use of cDNA libraries In deciding what to include we endeavored to combine up to date versions of some of the most widely used protocols with some very useful newer techniques *cDNA Library Protocols* should therefore be especially useful to the investigator who is new to the use of cDNA libraries but should also be of value to the more experienced worker Chapters 1-5 concentrate on cDNA library construction and manipulation Chapters 6 and 7 describe means of cloning difficult to obtain ends of cDNAs Chapters 8-18 give various approaches to the screening of cDNA libraries and the remaining chapters present methods of analysis of cDNA clones including details of how to analyze cDNA sequence data and how to make use of the wealth of cDNA data emerging from the human genome project

Short Protocols in Molecular Biology Frederick M. Ausubel, 1995-10-06 A desktop companion to the three volume *Current Protocols in Molecular Biology* the recognized leader in bioscience laboratory manuals This edition contains over 220 protocols from leading laboratories worldwide All methods are lab tested and include step by step instructions equipment and materials necessary to successfully conduct an experiment

Current Protocols in Molecular Biology Frederick M. Ausubel, 1987

The Biological Rationale for Therapeutic Targeting of the Four Transmembrane Protein, Epithelial Membrane Protein 2 (EMP2) Kaori Shimazaki, 2007 Cambridge Scientific

Biochemistry Abstracts, 1989 **Genetics Abstracts**, 1989 **Biotechnology Research Abstracts**, 1989 Monthly Classified listing of references to worldwide articles dealing with all aspects of biotechnology Also includes books and conferences Each entry gives bibliographic information institutional address of author(s) and abstract Author and subject index

Microbiology Abstracts, 1989-07

Reviewing **Protocol For Transformation Storage Solution 2x Tss**: Unlocking the Spellbinding Force of Linguistics

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